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TITLE: **PURIFIED NITRIC OXIDE SYNTHASE**

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**TO ALL WHOM IT MAY CONCERN:**

Be it known that We, John P.N. Rosazza and Yijun Chen have invented certain new and useful improvements in PURIFIED NITRIC OXIDE SYNTHASE of which the following is a specification:

**CROSS-REFERENCE TO RELATED APPLICATION**

This application is a continuation-in-part of copending application Serial No. 08/675,821, filed July 5, 1996, the disclosure of which is hereby incorporated by reference.

***Background of the Invention***

***1. Field of the Invention***

The present invention is directed to a purified enzyme and, more particularly, to a novel, purified, constitutive mammalian nitric oxide synthase (NOS) that utilizes both L-arginine and arginine-rich peptides, oligopeptides (*e.g.*, bradykinin (BK)), and proteins as substrates.

## 2. *Description of Related Art*

Nitric oxide synthases (NOS, EC 1.14.23) are important enzymes, which convert L-arginine to L-citrulline and nitric oxide (NO). Nitric oxide is a very short-lived free radical, which is rapidly oxidized to nitrite (NO<sub>2</sub>) and nitrate (NO<sub>3</sub>) which are measured as the stable inactive end products of nitric oxide formation. The significance, however, lies in the fact that NO appears to play a pivotal role in a wide variety of physiological and pathological processes in mammals. These processes include vasodilation and regulation of normal vascular tone, inhibition of platelet aggregation, neuronal transmission, cytostasis, hypotension associated with endotoxic shock, inflammatory response-induced tissue injury, mutagenesis, and formation of carcinogenic N-nitrosamines (Nathan, *FASEB J.* 6:3051-3064 (1992); Kiechle et al., *Am. J. Clin. Pathol.* 100:567-575 (1993)). For example, it is well-known in the art to treat humans afflicted with angina distress and cardiovascular disease with nitroglycerin, which acts as a vasodilating agent. In the body, nitroglycerin is converted to nitric oxide (NO), which is the pharmacologically active metabolite. See, Palmer *et al.*, *Nature* 333:664-666 (1988). Thus, evidence that NO mediates functions as diverse as those which occur in the brain, the endothelium and the blood, has led to intense study into the biological roles of NO and the various distinct members of the NOS family. (See, *e.g.*, Marletta, *J. Biol. Chem.* 268:12231-12234 (1993); Knowles *et al.*, *Biochem. J.* 298:249-258 (1994)).

It is well known by those skilled in the art that multiple isoforms of the NOS enzyme exist and that they are generally classified into two broad categories: 1) constitutive and 2) inducible. These classes of NOS enzymes vary considerably in their size, amino acid

sequence, activity and regulation, and exhibit a number of substantial differences, indicating differences in their molecular structures. Increasingly diverse biological functions are being attributed to the NO formed by these three major known types of NOS (Nathan, *FASEB J.* 6:3051 (1992); Marletta, *J. Biol. Chem.* 268:12231 (1993); Knowles *et al.*, *Biochem. J.* 298:249 (1994); Griffith *et al.*, *Annu. Rev. Physiol.* 57:707 (1995)). For example, cells such as neurons and vascular endothelial cells contain constitutive NOS isotypes, while macrophages and vascular endothelial cells express an inducible NOS.

Several isoforms of NOS's from different mammalian tissues and cells have been purified and characterized; in brain (nNOS) by Bredt and Snyder, *Proc. Natl. Acad. Sci. USA* 87:682-685 (1990); in endothelial cells (eNOS) by Forstermann *et al.*, *Biochem. Pharmacol.* 42:1849-1857 (1991); in macrophages (iNOS) by Hibbs *et al.*, *Science* 235:473 (1987) and Stuehr *et al.*, *Proc. Natl. Acad. Sci. U. S. A.* 88:7773-7777 (1991); in hepatocytes by Knowles *et al.*, *Biochem. J.* 279:833-836 (1990); in vascular cells by Wood *et al.*, *Biochem. Biophys. Res. Comm.* 170:80-88; in neutrophils by Yui *et al.*, *J. Biol. Chem.* 266:12544-12547 (1991) and Yui *et al.*, *J. Biol. Chem.* 266:3369-3371 (1991); and in other tissues (see, *e.g.*, Hevel *et al.*, *J. Biol. Chem.* 266:22789-22791 (1991); Mayer *et al.*, *FEBS Lett.* 277:215-219 (1990); Schmidt *et al.*, *Proc. Natl. Acad. Sci. U. S. A.* 88:365-369 (1991); Ohshima *et al.*, *Biochem. Biophys. Res. Commun.* 183:238-244 (1992); Hiki *et al.*, *J. Biochem.* 111:556-558 (1992); Evans *et al.*, *Proc. Natl. Acad. Sci. U. S. A.* 89:5361-5365 (1992); Sherman *et al.*, *Biochemistry* 32:11600-11605 (1993)). US Patent No. 5,268,465 claims a cDNA molecule encoding all or a portion of a mammalian calmodulin-dependent nitric oxide synthase (nNOS), comprising between 12 and 4,000 nucleotides.

US Patent No. 5,468,630 claims an isolated nucleic acid molecule comprising the nucleic acid sequence encoding a human inducible nitric oxide synthase (iNOS) protein. US Patent No. 5,498,539 claims an isolated nucleic acid molecule comprising the nucleic acid sequence encoding a bovine endothelial nitric oxide synthase (eNOS) protein having amino acid or nucleic acid sequences set forth in the specification.

It is also known that small amounts of NO generated by a constitutive NOS appear to act as a messenger molecule by activating soluble guanylate cyclase and, thus, increasing intracellular guanosine, 3', 5'-cyclic monophosphate (cGMP) and the induction of biological responses that are dependent on cGMP as a secondary messenger. For example, through this mechanism, endothelial derived NO induces relaxation of vascular smooth muscle and is identified as endothelium derived relaxing factor (Palmer *et al.*, *Nature* 327:524-526 (1987) and Ignarro *et al.*, *Proc. Natl. Acad. Sci. USA* 84:9265-9269 (1987)). In addition, neuronal nitric oxide can act as a neuro-transmitter by activating guanylate cyclase with important functions in the central nervous system and autonomic nervous systems. Bredt and Snyder, *Proc. Natl. Acad. Sci. USA* 86:9030-9033 (1989) and Burnett *et al.*, *Science* 257:401-403 (1992). Moreover, various purified NOS enzymes have been identified as hemoproteins (Stuehr *et al.*, *J. Biol. Chem.* 267:20547-20550 (1992); White *et al.*, *Biochemistry* 31:6627-6631 (1992); McMillan *et al.*, *Proc. Natl. Acad. Sci. USA* 89:11141-11145 (1992)) and flavoproteins (Hevel *et al.*, *J. Biol. Chem.* 266:22789-22791 (1991); Bredt *et al.*, *J. Biol. Chem.* 267:10976-10981 (1992)).

Thus, the catalytic mechanisms of the NOS enzymes have also been the subject of great interest (see, *e.g.*, Marletta, *J. Biol. Chem.* 268:12231-12234 (1993)). Stable isotope

studies have shown when that when L-arginine is the substrate for the enzyme, NO derives from one of the two equivalent guanidino nitrogens on the arginine moiety (Ignarro *et al.*, 1987; Palmer *et al.*, 1988), and that di-oxygen is the source of the oxygen atoms incorporated into citrulline and NO (Kwon *et al.*, *J. Biol. Chem.* 265:13442-13445 (1990); Leone *et al.*, *J. Biol. Chem.* 266:23790-23795 (1991)). Moreover, NG-hydroxy-L-arginine has been demonstrated to be an oxidative intermediate in the catalytic process (Stuehr *et al.*, *J. Biol. Chem.* 266:6259-6263 (1991); Wallace *et al.*, *Biochem. Biophys. Res. Commun.* 176:528-534 (1991); Pufahl *et al.*, *Biochemistry* 31:6822-6828 (1992); Klatt *et al.*, *J. Biol. Chem.* 268:14781-14787 (1993)).

Co-factors involved in the conversion of L-arginine to L-citrulline and NO synthesis include tetrahydrobiopterin (BH<sub>4</sub>), flavin adenine nucleotide (FAD), the flavin-ribitol phosphate part of FAD (FMN) and the reduced form of nicotinamide adenine dinucleotide phosphate (NADPH). In addition, when the NOS enzyme has been derived from brain or endothelial cells, calcium and calmodulin are also required. (Bredt *et al.*, *Nature* 351:714-718 (1991); Lamas *et al.*, *Proc. Natl. Acad. Sci. U. S. A.* 89:6348-6352 (1992); Lyons *et al.*, *J. Biol. Chem.* 267:6370-6374 (1992); Lowenstein *et al.*, *Proc. Natl. Acad. Sci. U. S. A.* 89:6711-6715 (1992); Xie *et al.*, *Science* 256:225-228 (1992)). Calmodulin is a well-known protein binder for Ca<sup>2+</sup>, ubiquitously found in plant and animal cells. The Ca<sup>2+</sup>-calmodulin complex thus formed is known to bind to various target proteins in the cell, and thereby alter their activity.

Thus, it will be appreciated that nitric oxide has both normal physiologic intracellular and extracellular regulatory functions. However, excessive production of

nitric oxide is detrimental. For example, when vascular endothelial cells are stimulated to express a NOS enzyme by a bacterial endotoxin, such as for example bacterial lipopolysaccharide (LPS), and inflammatory cytokines are elevated, the excess amounts of nitric oxides that are produced contribute to the vascular collapse seen in sepsis. Busse and Mulsch, *FEBS Lett.* 265:133-136 (1990). It is also known that when vascular cells are stimulated to express a NOS enzyme by inflammatory cytokines, the excess amounts of nitric oxides cause massive dilation of blood vessels and sustained hypotension commonly encountered in septic shock, and contribute to the eventual vascular collapse seen in sepsis. *Id.* It is also known that overproduction of nitric oxide in the lungs stimulated by immune complexes directly damages the lung. Mulligan *et al.*, *J. Immunol.* 148:3086-3092 (1992). Induction of nitric oxide synthase in pancreatic islets impairs insulin secretion and contributes to the onset of juvenile diabetes. Corbett *et al.*, *J. Biol. Chem.* 266:21351 (1991).

Thus, it will be appreciated that there is a great need in the medical community for the ability to control and regulate specific forms of NOS, particularly given its role in maintaining normal blood pressure and the devastating effect of excess NO on the cardiovascular, gastrointestinal and respiratory systems in humans. Thus considerable research has been expended to discover inhibitors and regulators of NOS activity. However, until the present invention, researchers have limited their work almost exclusively to the premise that L-arginine is the most physiologically relevant substrate of NOS. The studies have extended no further than the fact that dipeptides, such as Arg-Arg and Arg-Phe are also oxidized by crude NOS preparations derived from cultured

endothelial cells and macrophages (Hecker *et al.*, *FEBS Lett.* 294:221 (1991); Hecker *et al.*, *Proc. Natl. Acad. Sci. USA* 87:8612 (1990); Hecker *et al.*, *J. Cardiovasc. Pharmacol.* 20:S139 (1992)). Thus, although there have been many publications directed to analyses of arginine analogs or derivatives which inhibit NOS activity by blocking the use of arginine as a substrate for NO synthesis, or to the cofactors necessary for the conversion of arginine, no one until the present inventors has considered peptide, oligopeptide or protein substrates for the NOS enzyme. Hence, given the expectation that natural or synthetic arginine-rich peptide, oligopeptide or protein antagonists can function as <sup>1</sup>NOS inhibitors, the present invention will greatly expand the number and types of inhibitors available for the regulation and control of NO production in the body. In addition, the use of a NOS to supply deficient individuals with NO-forming capability will be greatly enhanced by the purification of the nNOS-II class of enzymes. The present invention, therefore, will also provide many new ways to study the biological mechanisms involved in NO synthesis.

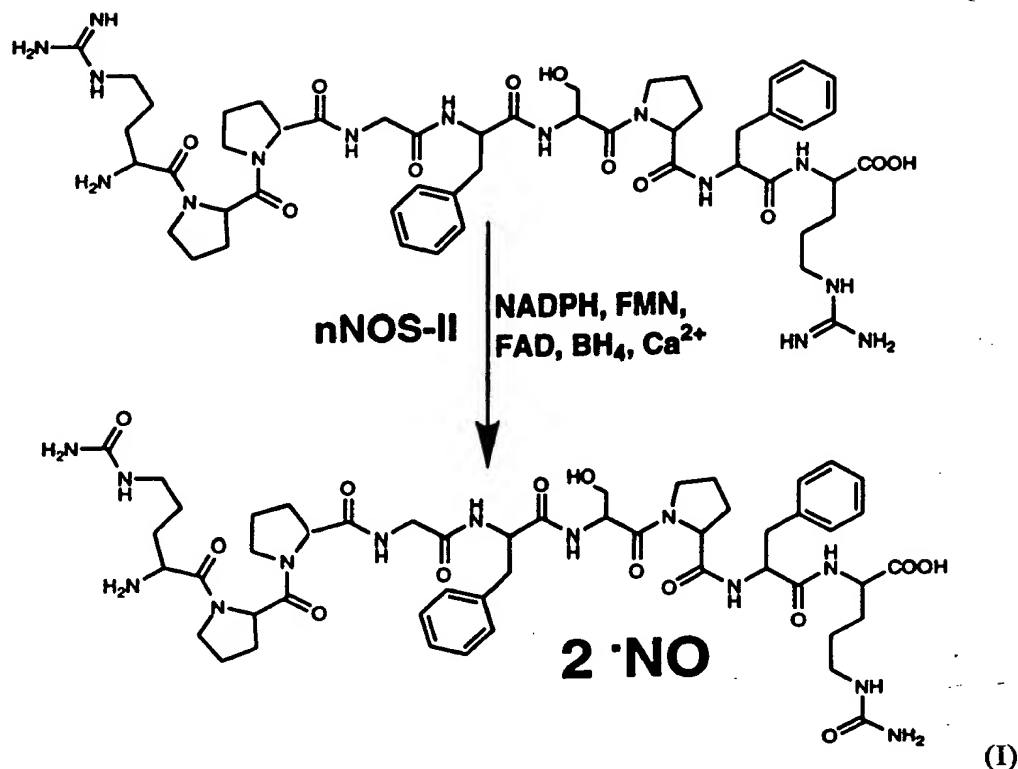
### ***Summary of the Invention***

The present invention relates to a novel constitutive nitric oxide synthase, referred to hereinafter as nNOS-II, that has been purified from mammalian neural tissue, and which is capable of utilizing arginine-rich peptides, polypeptides and proteins (*e.g.*, bradykinin), as well as L-arginine, as substrates. Unlike previously defined NOS isoforms, nNOS-II is unique in that it is calmodulin-dependent with L-arginine as substrate, but calmodulin-independent with an arginine-rich polypeptide, such as bradykinin (BK), as substrate. When BK is the substrate, both the N- and the C-terminal arginines of the oligopeptide are

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oxidized to citrullines by nNOS-II. See Equation I.

Equation I.



Moreover, with BK as substrate, NOS activity is competitively inhibited by N<sup>G</sup>-methyl-, N<sup>G</sup>-nitro-L-arginines, and BK receptor-antagonists, including oligopeptide BK receptor-antagonists.

More particularly, the present invention relates to a substantially purified nNOS-II protein.

In another embodiment, the present invention relates to a mammalian brain-derived nitric oxide synthase protein purified to an activity at least 6,360-fold, said protein having a denatured molecular mass as determined by sodium dodecyl sulfate polyacrylamide gel



electrophoresis under reducing conditions of about 105 kD, and a native homodimeric molecular mass as determined by gel filtration of about 230 kD, requiring NADPH, FAD, FMN,  $\text{Ca}^{2+}$  and tetrahydrobiopterin cofactors for the production of nitric oxide either from L-arginine, or an analog or derivative thereof, or from an arginine-rich peptide, oligopeptide, or protein substrate. As used herein, the term "peptide" is arbitrarily defined as a peptide chain having a single peptide bond, "oligopeptide" is a peptide chain having from 2 to 14 peptide bonds, inclusive, and "protein" is a peptide chain having 15 or more peptide bonds. "Polypeptide" is intended to be generic to "oligopeptide" and "protein."

"Arginine-rich" means that the peptide, oligopeptide, or protein has at least one sterically accessible arginine moiety.

In another embodiment, the present invention relates to a method of regulating or controlling nitric oxide production in a mammalian subject comprising administering to the mammal a nitric oxide-regulating amount of a peptide, oligopeptide, or protein inhibitor of nitric oxide synthase, preferably nNOS-II. As used herein, "mammalian subject" is intended to include human subjects.

In one embodiment, the present invention relates to a method of reducing the rate of nitric oxide production in a mammalian subject comprising administering to the mammal a nitric oxide inhibiting amount of a peptide, oligopeptide or protein inhibitor of nitric oxide synthase, preferably nNOS-II. "Reducing the rate" is intended to include a reduction to zero, in which case the reduction would be understood to include, not only a decrease in the rate of NO production or of the quantity of NO produced, but also a prevention of excess NO production.

In another embodiment, the present invention relates to a method of enhancing the rate of nitric oxide production in a mammalian subject comprising administering to the mammal a nitric oxide enhancing amount of nitric oxide synthase, preferably nNOS-II. "Enhancing the rate" is intended to include a stimulating, inducing or causing production of NO in a NO-deficient or NO-defective subject, in which case the enhancement would be understood to include stimulation, induction or initialization of NO production in such subjects or individuals, as well as increasing the rate of NO production or the quantity of NO produced.

In still another embodiment, the present invention relates to a method of preventing or treating a nitric oxide-mediated disease or condition in a mammalian subject comprising administering to the subject in need of such prevention or treatment a therapeutically effective amount of a peptide, oligopeptide, or protein inhibitor of nitric oxide synthase, preferably nNOS-II.

Additional objects, advantages and novel features of the invention will be set forth in part in the description which follows, and in part will become apparent to those skilled in the art on examination of the following, or may be learned by practice of the invention.

### ***Brief Description of the Drawings***

**Fig. 1** depicts a DEAE-agarose column elution profile for both NOS enzymes from rat cerebellum. L-arginine (○) NOS activity (calmodulin dependent), and BK (●) NOS

activity (calmodulin independent). Protein absorbance at 280 nm ( $\square$ ) from rat cerebellum was used to monitor the eluting process.

**Fig. 2** depicts an 8%, silver-stained SDS-polyacrylamide gel showing: *Lane 1*, cytosol (10  $\mu$ g protein); *Lane 2*, eluate from first DEAE-column (10  $\mu$ g); *Lane 3*, NADPH eluate from 2',5'-ADP-agarose column (2  $\mu$ g protein); *Lane 4*, eluate from second DEAE-agarose column containing purified enzyme (0.5  $\mu$ g protein); *Lane 5*, control with sample buffer; *Lane 6*, molecular weight markers (Bio-Rad) of myosin (208,000), b-galactosidase (115,000) phosphorylase B (107,000), bovine serum albumin (79,500), ovalbumin (49,500), carbonic anhydrase (34,800), soybean trypsin (28,300).

**Fig. 3** depicts HPLC chromatograms of nNOS-II reactions with BK or  $\text{Cit}^1$ -BK. Sensitivities of the regions at 20-30 minutes are expanded 3-fold. Fig. 3A is a chromatogram of the enzymatic reaction in which BK is the substrate. Fig. 3B is a chromatogram of the enzymatic reaction in which  $\text{Cit}^1$ -BK is the substrate.

**Fig. 4** depicts double reciprocal plots of the inhibition of NOS-II by [N-adamantaneacetyl-D-Arg<sup>0</sup>,Hyp<sup>3</sup>,Thi<sup>5,8</sup>,D-Phe<sup>7</sup>]-BK in the presence of BK. The inhibition experiments were carried out the same as for standard enzyme assays under initial velocity conditions. Concentrations of [N-adamantaneacetyl-D-Arg<sup>0</sup>,Hyp<sup>3</sup>,Thi<sup>5,8</sup>,D-Phe<sup>7</sup>]-BK were *closed circles*, 0  $\mu$ M; *open circles*, 0.625  $\mu$ M; *closed squares*, 1.25  $\mu$ M; *open squares*, 2.5  $\mu$ M; *closed triangles*, 5  $\mu$ M; *open triangles*, 10  $\mu$ M. The values are the mean of three measurements.

**Fig. 5** depicts DEAE-agarose column elution profile for NOSs and protein (absorbance at 280 nm.) from rat cerebellum. L-arginine (O) NOS activity (calmodulin dependent) and BK (●) NOS activity (calmodulin independent).

**Fig. 6** depicts an 8%, silver-stained SDS-polyacrylamide gel showing: Lane 1, cytosol (10 µg protein); Lane 2, eluate from first DEAE-agarose column (10 µg); Lane 3, NADPH eluate from 2'-5'-ADP-agarose column (2 µg protein); Lane 4, eluate from second DEAE-agarose column containing purified enzyme (0.5 µg protein); Lane 5, control with sample buffer; Lane 6, molecular weight markers (Bio-Rad) of myosin (208,000), b-galactosidase (115,000) phosphorylase B (107,000), bovine serum albumin (79,500), ovalbumin (49,500), carbonic anhydrase (34,800), soybean trypsin (28,300).

**Fig. 7** depicts HPLC chromatograms of nNOS-II reactions with BK or Cit<sup>1</sup>-BK. Sensitivities of the regions at 20-30 minutes are expanded 3-fold. (A) Chromatogram of enzymatic reaction with BK as substrate. (B) Chromatogram of enzymatic reaction with Cit<sup>1</sup>-BK as substrate.

**Fig. 8** depicts the oxidation of bradykinin (BK) to Cit<sup>1</sup>.Cit<sup>9</sup>-BK and nitric oxide (NO) by nNOS-II.

### ***Description of the Preferred Embodiments***

The present invention is directed to a novel constitutive mammalian nitric oxide synthase (NOS) that utilizes arginine-rich peptides, oligopeptides (*e.g.*, bradykinin), and proteins, as well as L-arginine, as substrates. The present invention was begun following

an earlier discovery by the inventors of a novel bacterial NOS that utilized a range of arginine-containing polypeptides, including BK and related BK analogs ranging from 6-mer to 11-mer, to produce nitric oxide. (See, Chen and Rosazza, *Biochem. Biophys. Res. Commun.* 203:1251 (1994) and Chen and Rosazza, *J. Bacteriol.* 177:5122 (1995)).

Although increasingly diverse biological functions are being attributed to NO formed by three major known types of NOS (Nathan, *FASEB J.* 6:3051-3064 (1992); Marletta, *J. Biol. Chem.* 268:12231-12234 (1993); Knowles *et al.*, *Biochem. J.* 298:249-258 (1994); Griffith *et al.*, *Annu. Rev. Physiol.* 57:707-736 (1995)), there are no known published reports of arginine-containing oligopeptides, polypeptides or proteins acting as a substrate for any NOS enzyme. To the contrary, throughout the prior art, L-arginine is currently recognized as the only physiologically relevant NOS-substrate.

The present inventors found that upon purification of crude supernatant preparations from rat cerebellum using weak anion exchange chromatography (DEAE-agarose), the elution profile reproducibly produces two calmodulin-dependent neuronal nitric oxide synthase (nNOS) peaks with L-arginine as substrate. Both peaks display the characteristic calmodulin-dependent nNOS activity using L-arginine as substrate, as described by Bredt *et al.*, *PNAS* 87:682(1990). However, the novel enzyme of the second peak (fractions 15-21), which has not been studied in the prior art, further displays unique calmodulin-independent nNOS activity in the presence of an arginine-rich polypeptide substrate, such as BK. (NOS is measured by the conversion of oxyhemoglobin to methemoglobin.) This novel nitric oxide synthase has been purified to afford a 6,360-fold purified enzyme preparation, and designated nNOS-II. When a species of nNOS-II acts upon a particular substrate, such as BK, the enzyme name may further denominate the

substrate, *e.g.*, nNOS<sub>BK</sub>. It should be emphasized, however, that BK is only one of many peptide, oligopeptide or protein substrates for nNOS-II.

Purified nNOS-II has a  $M_r$  of 105 kD by SDS-PAGE analysis and an apparent native  $M_r$  of 230 kD by gel filtration, indicating that the enzyme is a homodimeric protein. By comparison, the previously described, purified nNOS enzyme migrates as a single 160 kD band on SDS-PAGE, and the native enzyme appears to be a monomer. However, both enzymes require the presence of NADPH, FAD, FMN,  $Ca^{2+}$ , and tetrahydrobiopterin cofactors for substrate oxidation to occur.

When calmodulin is present with the necessary cofactors, nNOS-II also oxidizes L-arginine, but at a  $K_M$  of 10.6  $\mu M$ , slightly higher than the previously reported  $K_M$  of 2  $\mu M$  for nNOS. ( $K_M$  is the substrate concentration that allows the reaction to proceed at one-half its maximum rate.) Moreover, with an L-arginine substrate, the rate of reaction,  $V_{max}$ , value displayed by nNOS-II is 0.85  $\mu mol/min/mg$  protein, while that reported for the nNOS of the prior art is 0.96  $\mu mol/min/mg$  protein. ( $V_{max}$  refers to the rate of the enzyme reaction, depending only upon how rapidly the substrate molecule can be processed. This rate when divided by the enzyme concentration provides the turnover number.)

Finally, numerous reports describe the abolishment of nNOS activity (as defined by Bredt and Snyder) by the addition of quantities of L-arginine analog inhibitors. However, in marked contrast, with nNOS-II, using BK as substrate,  $N^G$ -methyl-L-arginine(L-NMA) and  $N^G$ -nitro-L-arginine(L-NNA) were found to be reversible competitive inhibitors with apparent  $K_i$  values of 8.6  $\mu M$  and 23.8  $\mu M$ , respectively. These values are significantly

different for the reported  $K_i$  values for L-NMA of 1.4  $\mu\text{M}$  and for L-NNA of 4.4  $\mu\text{M}$  for nNOS with L-arginine as substrate.

Thus, the present invention comprises a novel isoform of nitric oxide synthase, herein designated as nNOS-II, which can be readily distinguished from all previously reported NOS species, including nNOS. The broader implications of this discovery include the future identification of a new class of native, recombinant, or synthetic peptides that will function as NOS inhibitors for the modulation of cardiovascular, gastrointestinal, or bronchial activities, for contraceptive control, for the management of opioid withdrawal or cocaine-induced toxicity, or for the prevention or treatment of certain nitric oxide-mediated pathogenic conditions, such as ischemic stroke, diabetes, systemic hypotension, multiple sclerosis, Huntington's disease, Parkinson's disease, Alzheimer's disease, and the like.

It will be understood by those skilled in the art that the present invention is not limited to the use of any specific peptide, oligopeptide or protein as the substrate for the nNOS-II, although preferred substrates will be arginine-rich peptides, oligopeptides or proteins. More preferrably, the tertiary structure of such a peptide, oligopeptide or protein substrate will be such that one or more arginine groups are available to the NOS enzyme. It is most preferred that the substrate be an oligopeptide of manageable size, such as from 6 to 11 amino acid residues, wherein both the  $\alpha$  and  $\omega$  amino acids are arginine.

Exemplary peptides, oligopeptides and proteins upon which nNOS-II have been shown to be active are set forth in Table 1:

**Table 1: Arginine-Containing Peptides or Oligopeptides as Substrates for nNOS-II<sup>a</sup>**

Peptide	Amino Acid Sequence	Relative Activity (%)
L-arginine	N/A <sup>b</sup>	100 <sup>c</sup>
Poly-arginine (M <sub>r</sub> 5,000)	N/A	30
BK	Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg (SEQ ID NO:1)	125
Des-Arg <sup>1</sup> -BK	Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg (SEQ ID NO:2)	94
Des-Arg <sup>9</sup> -BK	Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe (SEQ ID NO:3)	180
BK fragment 1-7	Arg-Pro-Pro-Gly-Phe-Ser-Pro (SEQ ID NO:4)	80
BK fragment 1-5	Arg-Pro-Pro-Gly-Phe (SEQ ID NO:5)	61
BK fragment 2-7	✗ Pro-Pro-Gly-Phe-Ser-Pro (SEQ ID NO:6)	0
[Lys <sup>1</sup> ]-BK	Lys-Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg (SEQ ID NO:7)	113
Lys-BK	Lys-Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg (SEQ ID NO:8)	116
Ile-Ser-BK	Ile-Ser-Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg (SEQ ID NO:9)	110
Met-Lys-BK	✗ Met-Lys-Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg (SEQ ID NO:10)	0

<sup>a</sup> Standard enzymatic assay was carried out to measure initial velocity of the enzymatic reaction. Purified nNOS-II 0.2 µg (specific activity for BK, 280 nmole NO formed/min/mg protein) was used for the assays. Substrate concentrations were 100 µM except for poly-arginine used with 5 µg/ml.

<sup>b</sup> Not applicable.

<sup>c</sup> Activity for arginine is expressed as 100%. The values are the means of three measurements.

This list, however, is intended to be merely exemplary, not limiting in scope.



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A model substrate of nNOS-II is the nonapeptide bradykinin (Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg) (SEQ ID NO:1). BK was selected as a convenient and available model oligopeptide substrate for displaying the catalytic properties of nNOS-II because:

- it contains positionally different N- and C-terminal arginine residues;
- its recognized physiological role in both cardiovascular and central nervous systems has been well characterized;
- it displays physiological and pharmacological effects similar to those ascribed to NO; and
- it has been linked as an apparent mediator of NOS activity (Regoli *et al.*, *Pharmacol. Rev.* 32:1-46 (1980); Bhoola, *Pharmacol. Rev.* 44:1-58 (1992)).

In addition, BK is of ideal size (containing 9 amino acid residues) for conversion by the nNOS-II enzyme.

In a preferred embodiment, the present invention provides substantially pure nNOS-II protein. One specific enzyme herein provided in purified form is nNOS-II. When used with regard to nNOS-II of the present invention, the terms "protein" and "enzyme" are used interchangeably, even though technically an enzyme is a specific subset of the category "protein." Moreover, as used herein, a protein is said to be "highly purified" or "substantially pure" if the specific activity of the protein cannot be significantly increased by further purification, and if the specific activity is greater than that found in whole cell extracts containing the protein.

Any eukaryotic organism can be used as a source of nNOS-II, or the genes encoding same, as long as the source organism naturally contains the enzyme or its equivalent. As used herein, "source organism" refers to the original organism from which the amino acid

or DNA sequence is derived, regardless of the organism the enzyme expressed in or ultimately isolated from. For example, a human is said to be the "source organism" of nNOS-II expressed by a bacterial expression system as long as the amino acid sequence is that of human nNOS-II. The most preferred source organism is mammalian.

A variety of methodologies known in the art can be utilized to obtain the nNOS-II proteins of the present invention. In one embodiment, the enzyme is purified from tissues or cells which naturally produce it, such as rat cerebellum. One skilled in the art can readily follow known methods for isolating proteins in order to obtain the nNOS-II proteins. These include, but are not limited to, immunochromatography, size-exclusion chromatography, ion-exchange chromatography, affinity chromatography, HPLC, and the methods set forth by example in the present disclosure. One skilled in the art can readily adapt known purification schemes to delete certain steps or to incorporate additional purification procedures.

In a preferred embodiment of the invention, nNOS-II may be purified using column chromatography. Specifically, it has been found that greater than one-thousand-fold purification can be achieved using an affinity chromatography column. Since NADPH is a necessary cofactor for enzyme activity, if one employs a solid matrix containing an NADPH moiety or an NADPH analog, such as dextran blue, or 2',5'-ADP agarose or 2',5'-ADP sepharose, then the NOS of the present invention will bind to the matrix. It can be eluted using a soluble form of NADPH or an analog thereof at a concentration of about 1 to about 10 mM. It is desirable that the preparation which is applied to the affinity chromatography column first be partially purified on an ion exchange column, such as, diethylaminoethyl (DEAE) agarose.

Other ion exchange columns known in the art can also be used. The NOS of the present invention binds to DEAE-agarose and can be eluted with a sodium chloride gradient. The greatest peak of nNOS-II activity elutes with between about 150 mM and about 210 mM sodium chloride. A combination of these three column chromatography processes on a cleared brain homogenate will result in a homogeneous preparation, as can be demonstrated by silver staining of an SDS/PAGE-separated sample, or by Western blotting.

In another embodiment, the enzyme may be purified from cells which have been altered to express the desired protein. As used herein, a cell is said to be "altered to express a desired protein" when the cell, through genetic manipulation, is made to produce a protein which it normally does not produce, or which the cell normally produces at low levels. One skilled in the art can readily adapt procedures for introducing and expressing either genomic or cDNA sequences into either eukaryotic or prokaryotic cells, in order to generate a cell which produces the desired nNOS-II protein.

The present invention further encompasses the expression of the nNOS-II proteins (or a functional derivative thereof) in either prokaryotic or eukaryotic cells. A "functional derivative" of a sequence, either protein or nucleic acid, is a molecule that possesses a biological activity (either functional or structural) that is substantially similar to a biological activity of the protein or nucleic acid sequence. A functional derivative of a protein may or may not contain post-translational modifications such as covalently linked carbohydrate, depending on the necessity of such modifications for the performance of a specific function. The term "functional derivative" is intended to include the "fragments," "segments," "variants," "analogs," or "chemical derivatives" of a molecule.

As used herein, a molecule is said to be a "chemical derivative" of another molecule when it contains additional chemical moieties not normally a part of the molecule. Such moieties may improve the molecule's solubility, absorption, biological half life, and the like. The moieties may alternatively decrease the toxicity of the molecule, eliminate or attenuate any undesirable side effect of the molecule, and the like. Moieties capable of mediating such effects are disclosed in *Remington's Pharmaceutical Sciences* (1980). Procedures for coupling such moieties to a molecule are well known in the art.

A "variant" or "allelic or species variant" of a protein or nucleic acid is meant to refer to a molecule substantially similar in structure and biological activity to either the protein or nucleic acid. Thus, provided that two molecules possess a common activity and may substitute for each other, they are considered variants as that term is used herein even if the composition or secondary, tertiary, or quaternary structure of one of the molecules is not identical to that found in the other, or if the amino acid or nucleotide sequence is not identical.

Preferred prokaryotic hosts include bacteria such as *E. coli*, *Bacillus*, *Streptomyces*, *Pseudomonas*, *Salmonella*, *Serratia*, etc. Under such conditions, the nNOS-II will not be glycosylated. The prokaryotic host must be compatible with the replicon and control sequences in the expression plasmid.

However, prokaryotic systems may not prove efficacious for the expression of all proteins. While prokaryotic expression systems, *e.g.*, pET3c, have been used to express high molecular weight proteins, such as a biologically active (molecular weight ( $M_r$ ) ~118 kDa) FGF-1: $\beta$ -galactosidase chimera (Shi et al., submitted to *J. Biol. Chem.*, 1996), successful folding and disulfide bond formation may be difficult to accomplish in bacteria.

Nevertheless, to express nNOS-II (or a functional derivative thereof) in a prokaryotic cell, it is necessary to operably link the nNOS-II coding sequence to a functional prokaryotic promoter. Such promoters may be either constitutive or, more preferably, regulatable (*i.e.*, inducible or derepressible). Examples of constitutive promoters include the *int* promoter of bacteriophage  $\lambda$ , the *bla* promoter of the  $\beta$ -lactamase gene sequence of pBR322, and the CAT promoter of the chloramphenicol acetyl transferase gene sequence of pPR325, etc. Examples of inducible prokaryotic promoters include the major right and left promoters of bacteriophage  $\lambda$  ( $P_L$  and  $P_R$ ), the *trp*, *recA*, *lacZ*, *lacI*, and *gal* promoters of *E. coli*, the  $\alpha$ -amylase (Ulmanen *et al.*, *J. Bacteriol.* 162:176-182 (1985)) and the  $\zeta$ -28-specific promoters of *B. subtilis* (Gilman *et al.*, *Gene sequence* 32:11-20 (1984)), the promoters of the bacteriophages of *Bacillus* (Gryczan, In: *The Molecular Biology of the Bacilli*, Academic Press, Inc., NY (1982)), and *Streptomyces* promoters (Ward *et al.*, *Mol. Gen. Genet.* 203:468-478 (1986)). See also reviews by Glick (*J. Ind. Microbiol.* 1:277-282 (1987)); Cenatiempo (*Biochimie* 68:505-516 (1986)); and Gottesman (*Ann. Rev. Genet.* 18:415-442 (1984)).

Proper expression in a prokaryotic cell also requires the presence of a ribosome binding site upstream of the gene sequence-encoding sequence. Such ribosome binding sites are disclosed, for example, by Gold *et al.* (*Ann. Rev. Microbiol.* 35:365-404 (1981)).

Preferred eukaryotic hosts include yeast, fungi, insect cells, mammalian cells, either *in vivo* or in tissue culture. Mammalian cells which may be useful as hosts include HeLa cells, cells of fibroblast origin such as VERO or CHO-K1, or cells of lymphoid origin, such as the hybridoma SP2/O-AG14 or the myeloma P3x63Sg8, and their derivatives. Preferred

mammalian host cells include SP2/0 and J558L, as well as neuroblastoma cell lines such as IMR 332 that may provide better capacities for correct post-translational processing.

For a mammalian host, several possible vector systems are available for the expression of nNOS-II. A wide variety of transcriptional and translational regulatory sequences may be employed, depending upon the nature of the host. The transcriptional and translational regulatory signals may be derived from viral sources, such as adenovirus, bovine papilloma virus, Simian virus, or the like, where the regulatory signals are associated with a particular gene sequence which has a high level of expression. Alternatively, promoters from mammalian expression products, such as actin, collagen, myosin, etc., may be employed. Transcriptional initiation regulatory signals may be selected which allow for repression or activation, so that expression of the gene sequences can be modulated. Of interest are regulatory signals which are temperature-sensitive so that by varying the temperature, expression can be repressed or initiated, or are subject to chemical (such as metabolite) regulation.

Yeast expression systems can also carry out post-translational peptide modifications. A number of recombinant DNA strategies exist which utilize strong promoter sequences and high copy number of plasmids which can be utilized for production of the desired proteins in yeast. Yeast recognizes leader sequences on cloned mammalian gene sequence products and secretes peptides bearing leader sequences (*i.e.*, pre-peptides). Any of a series of yeast gene sequence expression systems incorporating promoter and termination elements from the actively expressed gene sequences coding for glycolytic enzymes produced in large quantities when yeast are grown in mediums rich in glucose can be utilized. Known glycolytic gene sequences can also provide very efficient

transcriptional control signals. For example, the promoter and terminator signals of the phosphoglycerate kinase gene sequence can be utilized.

The more preferred host for a protein the size of nNOS-II is insect cells, for example the *Drosophila* larvae. Using insect cells as hosts, the *Drosophila* alcohol dehydrogenase promoter can be used (see, e.g., Rubin, G.M., *Science* 240:1453-1459 (1988)).

The baculovirus insect cell expression system is the most preferred system for expressing the soluble nNOS-II construct as a carboxy-terminal triple tandem myc-epitope repeat:glutathione-S-transferase (GST) fusion protein chimera, using conventional PCR methods (Zhan *et al.*, *J. Biol. Chem.* 269:20221-20224 (1994)). These include the use of recombinant circle PCR to synthesize the soluble nNOS-II construct, the preparation and expression of the recombinant virus, AcNPV-GsJ in Sf9 cells (Summers and Smith (1988) A Manual of Methods for Baculovirus Vectors and Insect Culture Procedures (Texas Experimental Station Bulletin #1555)), the use of GST affinity chromatography (Zhan *et al.*, 1994) and reversed phase or ion exchange HPLC to purify the recombinant protein from Sf9 cell lysates and Myc immunoblot analysis to monitor the purification and assess the purity of the nNOS-II protein.

The soluble nNOS-II construct may not only prove to be valuable for the baculovirus expression system, but also as a construct for the expression of a secreted and soluble nNOS-II enzyme in mammalian cells for implantation *in vivo*. Moreover, baculovirus vectors can be engineered to express large amounts of nNOS-II in insect cells (Jasny, *Science* 238:1653 (1987); Miller *et al.*, in *Genetic Engineering* (1986), Setlow *et al.*, eds., *Plenum*, Vol. 8, pp. 277-297).

As discussed above, expression of nNOS-II in eukaryotic hosts requires the use of eukaryotic regulatory regions. Such regions will, in general, include a promoter region sufficient to direct the initiation of RNA synthesis. Preferred eukaryotic promoters include: the promoter of the mouse metallothionein I gene sequence (Hamer *et al.*, *J. Mol. Appl. Gen.* 1:273-288 (1982)); the TK promoter of Herpes virus (McKnight, *Cell* 31:355-365 (1982)); the SV40 early promoter (Benoist *et al.*, *Nature (London)* 290:304-310 (1981)); the yeast *gal4* gene sequence promoter (Johnston *et al.*, *Proc. Natl. Acad. Sci. (USA)* 79:6971-6975 (1982); Silver *et al.*, *Proc. Natl. Acad. Sci. (USA)* 81:5951-5955 (1984)).

As is widely known, translation of eukaryotic mRNA is initiated at the codon which encodes the first methionine. For this reason, it is preferable to ensure that the linkage between a eukaryotic promoter and a DNA sequence which encodes nNOS-II (or a functional derivative thereof) does not contain any intervening codons which are capable of encoding a methionine (*i.e.*, AUG). The presence of such codons results either in a formation of a fusion protein (if the AUG codon is in the same reading frame as the nNOS-II coding sequence) or a frame-shift a frame-shift mutation (if the AUG codon is not in the same reading frame as the nNOS-II coding sequence).

The nNOS-II coding sequence and an operably linked promoter may be introduced into a recipient prokaryotic or eukaryotic cell either as a non-replicating DNA (or RNA) molecule, which may either be a linear molecule or, more preferably, a closed covalent circular molecule. Since such molecules are incapable of autonomous replication, the expression of the nNOS-II may occur through the transient expression of the introduced sequence. Alternatively, permanent expression may occur through the integration of the introduced sequence into the host chromosome.



In one embodiment, a vector is employed which is capable of integrating the desired gene sequences into the host cell chromosome. Cells which have stably integrated the introduced DNA into their chromosomes can be selected by also introducing one or more markers which allow for selection of host cells which contain the expression vector. The marker may provide for prototrophy to an auxotrophic host, biocide resistance, e.g., antibiotics, or heavy metals, such as copper, or the like. The selectable marker gene sequence can either be directly linked to the DNA gene sequences to be expressed, or introduced into the same cell by co-transfection. Additional elements may also be needed for optimal synthesis of single chain binding protein mRNA. These elements may include splice signals, as well as transcription promoters, enhancers, and termination signals. cDNA expression vectors incorporating such elements include those described by Okayama, H., *Molec. Cell. Biol.* 3:280 (1983).

In a preferred embodiment, the introduced sequence will be incorporated into a plasmid or viral vector capable of autonomous replication in the recipient host. Any of a wide variety of vectors may be employed for this purpose. Factors of importance in selecting a particular plasmid or viral vector include: the ease with which recipient cells that contain the vector may be recognized and selected from those recipient cells which do not contain the vector; the number of copies of the vector which are desired in a particular host; and whether it is desirable to be able to "shuttle" the vector between host cells of different species.

Preferred prokaryotic vectors include plasmids, such as those capable of replication in *E. coli* (such as, for example, pBR322, ColE1, pSC101, pACYC 184,  $\pi$ VX). Such plasmids are, for example, disclosed by Maniatis *et al.* (In: *Molecular Cloning, A*



transformed cells as such, or following the induction of these cells to differentiate (for example, by administration of bromodeoxyuracil to neuroblastoma cells or the like).

The nNOS-II proteins (or a functional derivatives thereof) of the present invention can be used in a variety of procedures and methods, such as for the generation of antibodies, for use in identifying pharmaceutical compositions, for studying DNA/protein interaction, and for examinining the mechanism of NO synthesis.

The substantially pure peptides of the present invention may also be administered to a mammal intravenously, intramuscularly, subcutaneously, enterally, topically or parenterally. A "substantially pure" or "highly purified" protein, as defined previously, is a protein preparation that is generally lacking in other cellular components with which it is normally associated *in vivo*.

When administering peptides by injection, the administration may be by continuous injections, or by single or multiple injections. The peptides are intended to be provided to a recipient mammal in a "pharmacologically or pharmaceutically acceptable form" in an amount sufficient to "therapeutically effective." A peptide is considered to be in "pharmaceutically or pharmacologically acceptable form" if its administration can be tolerated by a recipient patient. An amount is said to be "therapeutically effective" (an "effective amount") if the dosage, route of administration, etc., of the agent are sufficient to effect a response to nNOS-II. Thus, the present peptides may be used to induce, increase, enhance, control, regulate or modulate the effect of the nNOS-II protein, or the synthesis and expression of NO.

In another embodiment of the present invention, methods for inhibiting, decreasing or preventing the activity of the nNOS-II protein can be achieved by providing an agent

capable of binding to or inhibiting the enzyme (or a functional derivative thereof). Such agents include, but are not limited to: nNOS-II inhibitors and antagonists, antisense nNOS-II, the antibodies to nNOS-II (anti-nNOS-II), and the secondary or anti-peptide peptides of the present invention. By decreasing the activity of nNOS-II, the effect which expression of the peptide has on NO synthesis can be modified, regulated, controlled, inhibited or prevented.

In one example of the present invention, methods are presented for decreasing the activity of nNOS-II (or a functional derivative thereof) by means of an anti-sense strand of cDNA to disrupt the translation of the nNOS-II message. Specifically, a cell is modified using routine procedures such that it expresses an antisense message, a message which is complementary to the pseudogene message. By constitutively or inducibly expressing the antisense RNA, the translation of the nNOS-II mRNA can be regulated. Such antisense technology has been successfully applied to regulate the expression of poly(ADP-ribose) polymerase (*see, Ding et al., J. Biol. Chem.* 267 (1992)).

In the alternative, nNOS-II activity can be prevented or inhibited by binding the peptide, oligopeptide or protein substrate used by the enzyme, thus modifying the amount of NO can be synthesized by the enzyme. Examples of such nNOS inhibitors include peptides or petidomimetics of structures such as bradykinin B2 receptor antagonists that have demonstrated nNOS-II inhibition activity, or arginine derivatives, such as L-NNA or L-NMA. Not all bradykinin B2 receptor antagonists have nNOS-II inhibitory activity as demonstrated in the present invention; however, it is within the ordinary skill of one in the art using known techniques and the procedures herein disclosed to determine which peptides, oligopeptides or proteins are capable of nNOS-II inhibition.

On the other hand, methods for stimulating, increasing or enhancing the activity of the nNOS-II peptide can be achieved by providing an agent capable of modulating the synthesis of NO by nNOS-II (or a functional derivative thereof), or by inhibiting or preventing an inhibitory signal which would diminish or stop the activity of nNOS-II in the system. Such agents include, but are not limited to, the anti-antisense nNOS-II peptides. By enhancing the activity of nNOS-II, the effect which the enzyme has on NO synthesis can also be modified.

In yet another embodiment, nNOS-II (or a functional derivative or variant thereof) can be used to produce antibodies or hybridomas. One skilled in the art will recognize that if an antibody is desired that will bind to nNOS-II, such an antibody would be generated as described above and used as an immunogen. The resulting antibodies are screened for the ability to bind nNOS-II.

The antibodies utilized in the above methods can be monoclonal or polyclonal antibodies, as well fragments of these antibodies and humanized forms. Humanized forms of the antibodies of the present invention may be generated using one of the procedures known in the art such as chimerization or CDR grafting.

In general, techniques for preparing monoclonal antibodies are well known in the art (Campbell, "*Monoclonal Antibody Technology: Laboratory Techniques in Biochemistry and Molecular Biology*," Elsevier Science Publishers, Amsterdam, The Netherlands (1984); St. Groth *et al.*, *J. Immunol. Methods* 35:1-21 (1980). For example, in one embodiment an antibody capable of binding nNOS-II is generated by immunizing an

animal with a synthetic polypeptide whose sequence is obtained from a region of the nNOS-II protein.

Any animal (mouse, rabbit, etc.) which is known to produce antibodies can be utilized to produce antibodies with the desired specificity, although because of the large size of the nNOS-II molecule, the rabbit may be preferred. Methods for immunization are well known in the art. Such methods include subcutaneous or interperitoneal injection of the polypeptide. One skilled in the art will recognize that the amount of polypeptide used for immunization will vary based on the animal which is immunized, the antigenicity of the polypeptide and the site of injection.

The polypeptide may be modified or administered in an adjuvant in order to increase the peptide antigenicity. Methods of increasing the antigenicity of a polypeptide are well known in the art. Such procedures include coupling the antigen with a heterologous protein (such as globulin or  $\beta$ -galactosidase) or through the inclusion of an adjuvant during immunization.

For monoclonal antibodies, spleen cells from the immunized animals are removed, fused with myeloma cells, such as SP2/0-Ag14 myeloma cells, and allowed to become monoclonal antibody producing hybridoma cells. A hybridoma is an immortalized cell line which is capable of secreting a specific monoclonal antibody.

Any one of a number of methods well known in the art can be used to identify the hybridoma cell which produces an antibody with the desired characteristics. These include screening the hybridomas with an ELISA assay, western blot analysis, or radioimmunoassay (Lutz *et al.*, *Exp. Cell Res.* 175:109-124 (1988)).

Hybridomas secreting the desired antibodies are cloned and the class and subclass are determined using procedures known in the art (Campbell, *Monoclonal Antibody Technology: Laboratory Techniques in Biochemistry and Molecular Biology*, Elsevier Science Publishers, Amsterdam, The Netherlands (1984)).

For polyclonal antibodies, antibody containing antisera is isolated from the immunized animal and is screened for the presence of antibodies with the desired specificity using one of the above-described procedures.

Conditions for incubating an antibody with a test sample vary. Incubating conditions depend on the format employed in the assay, the detection methods employed the nature of the test sample, and the type and nature of the antibody used in the assay. One skilled in the art will recognize that any one of the commonly available immunological assay formats (such as, radioimmunoassays, enzyme-linked immunosorbent assays, diffusion based Ouchterlony, or rocket immunofluorescent assays, or the like) can readily be adapted to employ the antibodies of the present invention. Examples of such assays can be found in Chard, "*An Introduction to Radioimmunoassay and Related Techniques*" Elsevier Science Publishers, Amsterdam, The Netherlands (1986); Bullock *et al.*, "*Techniques in Immunocytochemistry*," Academic Press, Orlando, FL Vol. 1 (1982), Vol. 2 (1983), Vol. 3 (1985); Tijssen, "*Practice and Theory of Enzyme Immunoassays: Laboratory Techniques in Biochemistry and Molecular Biology*," Elsevier Science Publishers, Amsterdam, The Netherlands (1985).

The anti-nNOS-II antibody and nNOS-inhibitors are also effective when immobilized on a solid support. Examples of such solid supports include, but are not

limited to, plastics such as polycarbonate, complex carbohydrates such as agarose and sepharose, and acrylic resins, such as polyacrylamide and latex beads. Techniques for coupling antibodies to such solid supports are well known in the art (Weir *et al.*, "Handbook of Experimental Immunology" 4th Ed., Blackwell Scientific Publications, Oxford, England, Chapter 10 (1986), Jacoby *et al.*, *Meth. Enzym.* 34 Academic Press, N.Y. (1974).

Additionally, one or more of the antibodies used in the above described methods can be detectably labelled prior to use. Antibodies can be detectably labelled through the use of radioisotopes, affinity labels (such as, biotin, avidin, etc.), enzymatic labels (such as, horse radish peroxidase, alkaline phosphatase, etc.) fluorescent labels (such as, FITC or rhodamine, etc.), paramagnetic atoms, etc. Procedures for accomplishing such labelling are well-known in the art, for example, see Sternberger *et al.*, *J. Histochem. Cytochem.* 18:315 (1970), Bayer *et al.*, *Meth. Enzym.* 62:308 (1979), Engval *et al.*, *Immunol.* 109:129 (1972), Goding, *J. Immunol. Meth.* 13:215 (1976). The labeled antibodies of the present invention can be used for *in vitro*, *in vivo*, and *in situ* assays to identify cells or tissues which express a specific protein or enzyme.

In an embodiment of the above methods, the antibodies are labeled, such that a signal is produced when the antibody(s) bind to the same molecule. One such system is described in U.S. Patent No. 4,663,278.

The antibodies or antisense peptides of the present invention may be administered to a mammal intravenously, intramuscularly, subcutaneously, enterally, topically or



parenterally. When administering antibodies or peptides by injection, the administration may be by continuous injections, or by single or multiple injections.

The antibodies or antisense peptides of the present invention are intended to be provided to a recipient mammal in a "pharmaceutically acceptable form" in an amount sufficient to be "therapeutically effective" or an "effective amount". As above, an amount is said to be therapeutically effective (an effective amount), if the dosage, route of administration, etc. of the agent are sufficient to affect the response to nNOS-II. Thus, the present antibodies may either stimulate or enhance the activity of the nNOS-II protein, resulting in increased NO synthesis, or they may inhibit or prevent the nNOS-II conversion of the peptide, oligopeptide or protein substrate into NO. Or, secondary antibody(s) may be designed to affect the response to the nNOS-II per se, *i.e.*, an anti-antibody to nNOS-II. In the alternative, either an antibody or an anti-antibody may be designed to affect only the anti-sense strand of the molecule.

One skilled in the art can readily adapt currently available procedures to generate secondary antibody peptides capable of binding to a specific peptide sequence in order to generate rationally designed antipeptide peptides, for example see Hurby et al., Application of Synthetic Peptides: Antisense Peptides", In *Synthetic Peptides, A User's Guide*, Freeman, NY, pp. 289-307 (1992), and Kaspczak *et al.*, *Biochemistry* 28:9230-8 (1989). As used herein, an agent is said to be "rationally selected or designed" when the agent is chosen based on the configuration of the nNOS-II peptide.

To detect secondary antibodies, or in the alternative, the labelled primary antibody, labelling reagents may include, *e.g.*, chromophobic, enzymatic, or antibody binding

reagents which are capable of reacting with the labelled antibody. One skilled in the art will readily recognize that the disclosed antibodies of the present invention can readily be incorporated into one of the established kit formats which are well known in the art.

An antibody is said to be in "pharmaceutically or pharmacologically acceptable form" if its administration can be tolerated by a recipient patient. The antibodies of the present invention can be formulated according to known methods of preparing pharmaceutically useful compositions, whereby these materials, or their functional derivatives, are combined with a pharmaceutically acceptable carrier vehicle. Suitable vehicles and their formulation, inclusive of other human proteins, e.g., human serum albumin, are described, for example, in Remington's *Pharmaceutical Sciences*, 1980).

In order to form a pharmaceutically acceptable composition which is suitable for effective administration, such compositions will contain an effective amount of an antibody of the present invention together with a suitable amount of carrier. Such carriers include, but are not limited to saline, buffered saline, dextrose, water, glycerol, ethanol, and a combination thereof. The carrier composition may be sterile. The formulation should suit the mode of administration. In addition to carriers, the antibodies of the present invention may be supplied in humanized form.

Humanized antibodies may be produced, for example by replacing an immunogenic portion of an antibody with a corresponding, but non-immunogenic portion (*i.e.*, chimeric antibodies) (Robinson *et al.*, International Patent Publication PCT/US86/02269; Akira *et al.*, European Patent Application 184,187; Taniguchi, European Patent Application 171,496; Morrison *et al.*, European Patent Application 173,494; Neuberger *et al.*, PCT

Application WO 86/01533; Cabilly *et al.*, European Patent Application 125,023; Better *et al.*, *Science* 240:1041-1043 (1988); Liu *et al.*, *Proc. Natl. Acad. Sci. USA* 84:3439-3443 (1987); Liu *et al.*, *J. Immunol.* 139:3521-3526 (1987); Sun *et al.*, *Proc. Natl. Acad. Sci. USA* 84:214-218 (1987); Nishimura *et al.*, *Canc. Res.* 47:999-1005 (1987); Wood *et al.*, *Nature* 314:446-449 (1985)); Shaw *et al.*, *J. Natl. Cancer Inst.* 80:1553-1559 (1988).

The compositions of the present invention can also include minor amounts of wetting or emulsifying agents, or pH buffering agents. The composition can be a liquid solution, suspension, emulsion, tablet, pill, capsule, sustained release formulation or powder. The composition can be formulated as a suppository with traditional binders and carriers such as triglycerides. Oral formulations can include standard carriers such as pharmaceutically acceptable mannitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate, etc.

In a preferred embodiment of the present invention, the compositions are formulated in accordance with routine procedures for intravenous administration to a subject. Typically, such compositions are carried in a sterile isotonic aqueous buffer. As needed, a composition may include a solubilizing agent and a local anesthetic. Generally, the components are supplied separately or as a mixture in unit dosage form, such as a dry lyophilized powder in a sealed container with an indication of active agent. Where the composition is administered by infusion, it may be provided with an infusion container with a sterile pharmaceutically acceptable carrier. When the composition is administered by injection, an ampoule of sterile water or buffer may be included to be mixed prior to injection.

The therapeutic compositions may also be formulated in salt form.

Pharmaceutically acceptable salts include those formed with free amino groups, such as those derived from hydrochloric, phosphoric, acetic, oxalic and tartaric acids, or formed with free carboxyl groups such as those derived from sodium, potassium, ammonium, calcium, ferric hydroxides, isopropylamine, triethylamine, 2-ethylamino ethanol, histidine, procaine, etc.

The dosage of the administered agent will vary depending upon such factors as the patient's age, weight, height, sex, general medical condition, previous medical history, etc. In general, it is desirable to provide the recipient with a dosage of the antibody which is in the range of from about 1  $\mu\text{g}/\text{kg}$  to 10  $\text{mg}/\text{kg}$  (body weight of patient), although a lower or higher dosage may be administered. [Suitable ranges for intravenous administration is typically about 20-500  $\mu\text{g}$  of (active compound) per kilogram body weight] Effective doses may be extrapolated from dose-response curves derived from *in vitro* and *in vivo* animal model test systems.

Since highly purified proteins are now available, X-ray crystallography and NMR-imaging techniques can be used to identify the structure of the enzyme. Utilizing such information, computer modeling systems are now available that allows one to "rationally design" an agent capable of binding to a defined structure (Hodgson, *Biotechnology* 8:1245-1247 (1990)), Hodgson, *Biotechnology* 9:609-613 (1991)). As used herein, an agent is said to be "rationally designed" if it is selected based on a computer model of nNOS-II.

In another embodiment of the present invention, methods are provided for modulating the translation of RNA encoding nNOS-II protein in the cell. Specifically, said method comprises introducing into a cell a DNA sequence which is capable of transcribing RNA which is complimentary to the RNA encoding the nNOS-II protein. By introducing such a DNA sequence into a cell, antisense RNA will be produced which will hybridize and block the translation of the nNOS-II protein. Antisense cloning has been described by Rosenberg *et al.*, *Nature* 313:703-706 (1985), Preiss *et al.*, *Nature* 313:27-32 (1985), Melton, *Proc. Natl. Acad. Sci. USA* 82:144-148 (1985) and Kim *et al.*, *Cell* 42:129-138 (1985).

Transcription of the introduced DNA will result in multiple copies of antisense RNA which will be complimentary to the nNOS-II. By controlling the level of transcription of antisense RNA, and the tissue specificity of expression, one skilled in the art can regulate the level of activity of nNOS-II in specific cells within a patient.

In another embodiment of the present invention, kits are provided which contain the necessary reagents to carry out the previously described methods and assays.

All essential publications mentioned herein are hereby incorporated by reference.

In order that those skilled in the art can more fully understand this invention, the following examples are set forth. These examples are included solely for the purpose of illustration, and should not be considered as expressing limitations unless so set forth in the appended claims.

## *Examples*

In the following examples and protocols, restriction enzymes, ligase, labels, and all commercially available reagents were utilized in accordance with the manufacturer's recommendations. The cell and molecular and protein purification methods utilized in this application are established in the art and will not be described in detail. However, standard methods and techniques for isolation, purification, labeling, and the like, as well as the preparation of standard reagents may be performed essentially in accordance with *Molecular Cloning: A Laboratory Manual, 2nd ed.*, edited by Sambrook, Fritsch & Maniatis, Cold Spring Harbor Laboratory, 1989, and the revised third edition thereof, or as set forth in the literature references cited and incorporated herein. Methodologic details may be readily derived from the cited publications.

### *Example 1*

#### *Isolation, Purification and Characterization of nNOS-II*

Using the following materials and methods, substantially pure nNOS-II protein was prepared and purified from crude supernatant preparations from rat cerebellum, and characterized as containing calmodulin-independent NOS activity capable of catalyzing the oxidation of an arginine-rich nonapeptide, BK, to produce NO.

**Materials:** BK, [Thi<sup>5,8</sup>,D-Phe<sup>7</sup>]-BK, and [N-adamantaneacetyl-D-Arg<sup>0</sup>, Hyp<sup>3</sup>, Thi<sup>5,8</sup>, D-Phe<sup>7</sup>]-BK were purchased from American Peptide Co. (San Diego, CA). Cit<sup>1</sup>-BK was synthesized (University of Iowa, Protein Structure Facility) by solid phase synthesis using 2-chlorotrityl chloride resin (Barlos *et al.*, *Int. J. Peptide Protein Res.* 37:513-520 (1991)). The synthetic peptide Cit<sup>1</sup>-BK was greater than 98% pure by HPLC, and gave m/z

1061.3 by laser desorption mass spectrometry. (6R)-5,6,7,8-Tetrahydrobiopterin (BH<sub>4</sub>) was from Biochemical Research Inc. (Natick, MA). L-arginine, N<sup>G</sup>-methyl-L-arginine (L-NMA), N<sup>G</sup>-nitro-L-arginine (L-NNA), 2',5'-ADP-agarose and other reagents were purchased from Sigma Chemical Co. (St Louis, MO).

**Enzyme Purification:** Ten cerebella taken from Sprague Dailey rats (male, 250-350 g) were homogenized in 50 mL of ice-cold buffer A {10 mM Tris-HCl (pH 7.5) containing 1 mM DTT, 1 mM EDTA}. All subsequent purification procedures were carried out at 4°C. The homogenate was centrifuged at 100,000 x g for 120 min, and the supernatant was loaded onto a 25 mL DEAE-agarose column equilibrated with buffer A. The column was washed with 50 mL buffer A and eluted with a 200 mL linear gradient of 0-500 mM NaCl in buffer A. Fractions (4 mL) were collected and assayed for enzyme activity.

Elution profiles similar to those observed by Bredt *et al.* (*Proc. Natl Acad. Sci. USA* 87:682-685 (1990)) reproducibly gave two calmodulin-dependent NOS activity peaks with L-arginine as substrate (see Fig. 1, fractions 8-21 (○)). However, only one of these (Fig. 1, fractions 15-21 (●)) displayed calmodulin-independent NOS activity with BK as substrate. Protein absorbance at 280 nm (Fig. 1, □) from rat cerebellum was used to monitor the eluting process.

Fractions representing the active peak for BK from the DEAE-agarose column were pooled and concentrated to 15 mL by membrane ultrafiltration, and loaded onto a 5 mL, 2',5'-ADP-agarose column equilibrated with buffer B {10 mM Tris-HCl (pH 7.0) containing 1 mM DTT, 1 mM EDNA, and 10% glycerol}. The column was subsequently

washed with 20 mL of buffer B, 10 mL of 0.5 M NaCl in buffer B, 10 mL of 0.5 mM NADH in buffer B, 10 mL of 0.5 mM NADP<sup>+</sup> in buffer B, and 20 mL of buffer B. The NOS activity was finally eluted with 10 mL of 10 mM NADPH in buffer B.

The affinity eluate was concentrated to 1.5 mL and loaded onto a 5 mL DEAE-agarose column equilibrated with buffer B. The column was washed with 20 mL buffer B and eluted with a 100 mL linear gradient of 0-300 mM NaCl in buffer B. Fractions (2 mL) were collected and assayed for enzyme activity, and active fractions (8 mL) were pooled and concentrated to 0.2-0.5 mL with an Amicon PM-30 membrane to afford a 6,360-fold purified enzyme preparation. The thus-purified, novel constitutive neuronal NOS was designated as nNOS-II to differentiate it from the NOS isoform (nNOS) previously described by Bredt *et al.*, *supra* (1990).

The purification of nNOS-II is summarized in Table 2:

**Table 2. Purification of nNOS-II from rat cerebellum.\***

Step	Total Protein (μg)	Total Activity (nmol/min)	Specific Activity (nmol/min/mg protein)	Recovery (%)	Purification Factor (fold)
Cytosol	356,640	44.18	0.19	100	1
DEAE-agarose	69,200	27.20	0.39	61.6	2
ADP-agarose	66	19.10	289.1	43.2	1,522
DEAE-agarose	4	4.64	1,208.3	10.5	6,360

\* Values of final specific activities, recovery and fold-purification represent the means of three enzyme preparations. Enzyme activity was measured spectrophotometrically. BK (100 μM) was used for enzyme assay during enzyme purification, and enzymatic reactions were initiated by the addition of protein. Protein was determined using the Bradford protein assay.



**NOS Activity Assay:** NOS activity was determined spectrophotometrically by the rapid and quantitative oxidation of oxyhemoglobin to methemoglobin (Feelish *et al.*, *Eur. J. Pharmacol.* 139:19-30 (1987); Olken *et al.*, *Biochem. Biophys. Res. Commun.* 177:828-833 (1991)).

Like previously characterized nNOS from cerebellum, nNOS-II requires NADPH, FAD, FMN,  $\text{Ca}^{2+}$ , and  $\text{BH}_4$  with BK as substrate. Thus, reaction mixtures for NOS assays containing 50 mM Tris-HCl buffer (pH 7.5), were optimized to contain 4  $\mu\text{M}$  oxyhemoglobin, 100  $\mu\text{M}$  NADPH, 1 mM  $\text{CaCl}_2$ , 10  $\mu\text{M}$  FAD, 10  $\mu\text{M}$  FMN, 20  $\mu\text{M}$   $\text{BH}_4$ , 150  $\mu\text{M}$  DTT, 100  $\mu\text{M}$  BK or L-arginine and 0.1-3  $\mu\text{g}$  enzyme in a final volume of 0.5 mL.

nNOS-II has stabilities similar to nNOS. In addition, nNOS-II also oxidizes L-arginine, but *only* when calmodulin is added along with the other cofactors. Thus, calmodulin (10  $\mu\text{g/mL}$ ) was added to the reaction mixtures when L-arginine was used as substrate.

**Determinations of Molecular Weight:** The molecular mass of the denatured purified enzyme was determined by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) to be approximately 105 kD (*see*, Fig. 2). The apparent native molecular mass of the enzyme was approximately 230 kD by gel filtration, indicating that nNOS-II is a homodimeric protein.

The native molecular weight of the purified enzyme was estimated by analytical gel filtration chromatography carried out using an Alltech Macrosphere 150 column (7  $\mu\text{m}$ , 4.6 x 25 cm) and a mobile phase of 10 mM Tris-HCl buffer (pH 7.5) containing 1 mM DTT and 0.2 M NaCl, which was used to equilibrate the column and to elute protein samples at a

flow rate of 0.5 mL/min. Eluting protein peaks (retention volumes,  $R_v$ ) were monitored at 280 nm. Standard proteins ( $M_r$ ) used (as shown in Fig. 2) were bovine thyroglobulin (669,000,  $R_v$  1.83 mL), horse spleen apoferritin (443,000,  $R_v$  1.97 mL), sweet potato  $\beta$ -amylase (200,000,  $R_v$  2.28 mL), yeast alcohol dehydrogenase (150,000,  $R_v$  2.47 mL), and bovine serum albumin (66,000,  $R_v$  2.78 mL).

The peak ( $R_v$  2.25 mL) corresponding to the eluted enzyme (230,000) was collected and assayed for NOS activity.

**Kinetic Determinations:** Since BK contains both N- and C-terminal arginines, it was possible that BK could give rise to NO plus three different nonapeptide products including: Cit<sup>1</sup>-BK, Cit<sup>9</sup>-BK and Cit<sup>1</sup>,Cit<sup>9</sup>-BK. HPLC and amino acid sequencing were used to isolate and identify the nonapeptide product formed when nNOS-II oxidized BK. Kinetic experiments were conducted using the standard enzyme assay described above.

#### **a) Isolation of Enzymatic Products by High Performance Liquid**

**Chromatography (HPLC):** Enzymatic reaction mixtures contained 0.5  $\mu$ g purified enzyme, 50 mM Tris-HCl buffer (pH 7.5), 100  $\mu$ M NADPH, 20  $\mu$ M FAD, 20  $\mu$ M FMN, 1 mM CaCl<sub>2</sub>, 50  $\mu$ M tetrahydrobiopterin, and 100  $\mu$ M substrate in final volumes of 0.5 mL. Reaction mixtures were incubated for 120 min at 37°C, and transferred to microconcentrators ( $M_r$  cut-off 10,000, Bio-Rad) and centrifuged to remove the enzyme. Fractions through the concentrator membrane were subjected to HPLC analyses. HPLC was performed with a Shimadzu LC-10AD pump and a SPD-M6A photodiode array UV-Vis detector set at 214-219 nm. Samples of 15  $\mu$ L were resolved on a  $\mu$ Bondapak C18 column (Waters; 10 mm; 3.9 x 300 mm, inside diameter) preceded by a guard column (3.9

x 20 mm) at a flow rate of 1 mL/min. The mobile phase consisted of mixtures of: A. 0.1% trifluoroacetic acid in water; B. 0.095% trifluoroacetic acid in acetonitrile.

Elution was achieved with the following gradients: 0-15 min, 0-15% linear gradient of B; 15-35 min, 15-20% linear gradient of B; and 35-40 min, 20-0% linear gradient of B. Retention times for BK, Cit<sup>1</sup>-BK, and Cit<sup>1</sup>,Cit<sup>9</sup>-BK are 22.8 min, 24.3 min, and 26.3 min, respectively.

For amino acid sequencing, reaction mixture samples of 200 µL were injected, and peaks at 26.3 min were collected and concentrated. Incubations without substrate and without purified enzyme were used and analyzed as controls.

**b) Amino Acid Sequencing:** Amino acid sequencing was determined by automated microsequencing with Edman degradation reactions on a 475A Sequencer (Applied Biosystems, Inc.) in the University of Iowa, Protein Structure Facility. The enzymatic product (2 µg) from HPLC (*see*, Figs. 3A and B) was sequenced in duplicate to confirm the peptide sequence.

**c) Kinetic Data:** To determine  $K_M$  values for BK, Cit<sup>1</sup>-BK and L-arginine, the substrate concentrations used were 3.125, 6.25, 12.5, 25, and 50 µM. L-NMA and L-NNA, at concentrations of 0, 3.125, 6.25, 12.5, 25, and 50 µM, were used to measure  $K_i$  values in the presence of above concentrations of BK. *N*-adamantaneacetyl-[D-Arg<sup>0</sup>, Hyp<sup>3</sup>, Thi<sup>5,8</sup>, D-Phe<sup>7</sup>]-BK, at concentrations of 0, 0.625, 1.25, 2.5, 5, and 10 µM, was used to determine its  $K_i$  value in the presence of the concentrations of BK given above. Kinetic data were calculated by fitting experimental data to the EZ-FIT program (Perrella, *Anal. Biochem.* 174:437-447 (1988)).

The only peptide product obtained during the nNOS-II oxidation of BK was Cit<sup>1</sup>,Cit<sup>9</sup>-BK, where both N- and C-terminal BK-arginines were converted to their corresponding citrullines (Fig. 3A). The synthetic nonapeptide Cit<sup>1</sup>-BK in which N-terminal arginine was replaced with citrulline was also a substrate for nNOS-II, and it, too, gave Cit<sup>1</sup>,Cit<sup>9</sup>-BK (Fig. 3B). Difficulties encountered in synthesizing Cit<sup>9</sup>-BK precluded its use in the present investigations.

Apparent  $K_M$  values for BK and Cit<sup>1</sup>-BK were 8.5 and 6.2  $\mu$ M, respectively, while apparent  $V_{max}$  values for BK and Cit<sup>1</sup>-BK were 1.2 and 1.6  $\mu$ mol/min/mg protein, respectively. The kinetic results suggest that neither Cit<sup>1</sup>-BK nor Cit<sup>9</sup>-BK are likely to accumulate as products during the course of nNOS-II oxidations of BK.

By comparison, the apparent  $K_M$  of nNOS-II for L-arginine was 10.6  $\mu$ M, slightly higher than the reported value of 2  $\mu$ M for previously described nNOS (Bredt *et al.*, *Proc. Natl Acad. Sci. USA* 87:682-685 (1990)). With L-arginine as substrate, nNOS-II and nNOS display apparent  $V_{max}$  values of 0.85 and 0.96  $\mu$ mol/min/mg protein, respectively.

With nNOS-II and BK as substrate, typical NOS inhibitors, L-NMA and L-NNA, are reversible, competitive inhibitors with apparent  $K_i$  values of 8.6  $\mu$ M and 23.8  $\mu$ M, respectively. These  $K_i$  values are, however, significantly different from previously established  $K_i$  values for L-NMA of 1.4  $\mu$ M and L-NNA of 4.4  $\mu$ M with L-arginine as substrate for nNOS (Nathan, *FASEB J.* 6:3051-3064 (1992); Marletta, *J. Biol. Chem.* 268:12231-12234 (1993); Knowles *et al.*, *Biochem. J.* 298:249-258 (1994); Griffith *et al.*, *Annu. Rev. Physiol.* 57:707-736 (1995)).

In addition, known potent specific B2 receptor antagonists [*N*-adamantaneacetyl-D-Arg<sup>0</sup>, Hyp<sup>3</sup>, Thi<sup>5,8</sup>, D-Phe<sup>7</sup>]-BK and [Thi<sup>5,8</sup>, D-Phe<sup>7</sup>]-BK (Lammek *et al.*, *Peptides* 11:1041-1043 (1990); Lammek *et al.*, *J. Phar. Pharmacol.* 43:887-888 (1988); Austin *et al.*, *J. Physiol.* 478:351-356 (1994)) were examined for their possible effects with regard to nNOS-II activity. While [*N*-adamantaneacetyl-D-Arg<sup>0</sup>, Hyp<sup>3</sup>, Thi<sup>5,8</sup>, D-Phe<sup>7</sup>]-BK competitively inhibited nNOS-II activity on BK, with an apparent K<sub>i</sub> of 2.5 mM, [Thi<sup>5,8</sup>, D-Phe<sup>7</sup>]-BK had no effect on nNOS-II activity over a concentration range of 0.5 - 100 mM.

In view of these findings, it is believed that natural endogenous peptide substrates other than BK exist for nNOS-II. It is also believed that oligopeptide-utilizing endothelial and macrophage NOSs will be found. The broad implications of this discovery are that arginine-rich peptides of greater or lesser size than BK can serve as NOS substrates to form NO, and that natural or synthetic peptides, oligopeptides or proteins can function as NOS.

Although the present invention has been described with reference to the presently preferred embodiments, the skilled artisan will appreciate that various modifications, substitutions, omissions and changes may be made without departing from the spirit of the invention. Accordingly, it is intended that the scope of the present invention be limited only by the scope of the following claims, including equivalents thereof.

## *Example 2*

### *Comparison of Reaction Rates with Bradykinin Analogs*

Relative reaction rates were compared with a series of BK analogs. As expected, most of the BK analogs studied served as substrates for nNOS-II. Table 11 shows the relative activities of the arginine-containing peptides as substrates for nNOS-II. Different than that for NOS<sub>Noc</sub>, poly-arginine only showed 30% activity for nNOS-II comparing to that of arginine. However, nonapeptide BK showed more activity than arginine; Moreover, the octapeptide, des-Arg<sup>1</sup>-BK, in which the N-terminal arginine is removed had activity similar to that of arginine. Another octapeptide, des-Arg<sup>9</sup>-BK, in which the C-terminal arginine is removed had activity about one-fold higher than that of arginine. The heptapeptide BK fragment 1-7 decreased its activity by 30% while BK fragment 1-5 had about 50% activity of that for BK. When the N-terminal arginine was replaced by lysine, nonapeptide [Lys<sup>1</sup>]-BK had similar activity to that for BK, while BK with an additional N-terminal lysine, decapeptide Lys-BK, had almost the same activity. However, Lys-BK with an additional methionine, the undecapeptide Met-Lys-BK, showed no activity at all. Although the length of peptides, the position of arginine in the peptides, and the presence of certain amino acids such as lysine may affect the activities for nNOS-II as substrates, there is no clear relationship between the peptide length or the position of arginine and NOS activity. These preliminary results indicate that BK and analogs can directly serve as substrates for nNOS-II, although structure-activity relationship could not finally established based on this study.

Table 3. Arginine-Containing Peptides as Substrates for nNOS-II<sup>a</sup>

Peptide	Amino acid sequence	Relative activity (%)
L-Arginine	N/A <sup>b</sup>	100 <sup>c</sup>
Poly-arginine (M <sub>r</sub> 5,000)	N/A	30
1 BK	Arg <sup>1</sup> -Pro <sup>2</sup> -Pro <sup>3</sup> -Gly <sup>4</sup> -Phe <sup>5</sup> -Ser <sup>6</sup> -Pro <sup>7</sup> -Phe <sup>8</sup> -Arg <sup>9</sup>	125
2 Des-Arg1-BK	Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg	94
3 Des-Arg9-BK	Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe	180
4 BK fragment 1-7	Arg-Pro-Pro-Gly-Phe-Ser-Pro	80
5 BK fragment 1-5	Arg-Pro-Pro-Gly-Phe	61
<del>6</del> BK fragment 2-7	Pro-Pro-Gly-Phe-Ser-Pro	0
7 [Lys1]-BK	Lys-Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg	113
8 Lys-BK	Lys-Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg	116
9 Ile-Ser-BK	Ile-Ser-Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg	110
Met-Lys-BK	Met-Lys-Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg	0

<sup>a</sup> Standard enzyme assay was carried out to measure initial velocity of the enzymatic reaction. Purified nNOS-II 0.2 µg (specific activity for BK, 280 nmole NO formed/min/mg protein) was used for the assays. Substrate concentrations were 100 µM except for poly-arginine used with 5 µg/ml.

<sup>b</sup> Not applicable.

<sup>c</sup> Activity for arginine is expressed as 100%. The values are the means of three measurements.

### Example 3

The following is hereby expressly incorporated in its entirety by reference:

Chen, Yijun and Rosazza, John P.N., "Oligopeptides as Substrates and Inhibitors for a New Constitutive Nitric Oxide Synthase from Rat Cerebellum", *Biochemical and Biophysical Research Communications*, 224:303-308 (1996), Article No. 1025.

## Oligopeptides as Substrates and Inhibitors for a New Constitutive Nitric Oxide Synthase from Rat Cerebellum

### MATERIALS AND METHODS

**Materials.** BK, [Thi<sup>5,8</sup>.D-Phe<sup>7</sup>]-BK, and [N-adamantanecacetyl-D-Arg<sup>0</sup>. Hyp<sup>3</sup>. Thi<sup>5,8</sup>. D-Phe<sup>7</sup>]-BK were purchased from American Peptide Co. (San Diego, CA). Cit<sup>1</sup>-BK was synthesized (University of Iowa, Protein Structure Facility) by solid phase synthesis using 2-chlorotrityl chloride resin (9). The synthetic peptide Cit<sup>1</sup>-BK was greater than 98% pure by HPLC, and gave m/z 1061.3 by laser desorption mass spectrometry. (6R)-5,6,7,8-Tetrahydrobiopterin (BH<sub>4</sub>) was from Biochemical Research Inc. (Natick, MA). L-Arginine, N<sup>G</sup>-methyl-L-arginine (L-NMA), N<sup>G</sup>-nitro-L-arginine (L-NNA), 2',5'-ADP-agarose and other reagents were purchased from Sigma Chemical Co. (St Louis, MO).

**NOS activity assay.** NOS activity was determined spectrophotometrically by the rapid and quantitative oxidation of oxyhemoglobin to methemoglobin (10,11). Optimized reaction mixtures for NOS assays contained 50 mM Tris-



HCl buffer (pH 7.5), and were optimized to contain 4  $\mu$ M oxyhemoglobin, 100  $\mu$ M NADPH, 1 mM  $\text{CaCl}_2$ , 10  $\mu$ M FAD, 10  $\mu$ M FMN, 20  $\mu$ M BH<sub>4</sub>, 150  $\mu$ M DTT, 100  $\mu$ M BK or L-arginine and 0.1-3  $\mu$ g enzyme in a final volume of 0.5 ml. Calmodulin (10  $\mu$ g/ml) was added to the reaction mixtures when L-arginine was used as substrate.

**Enzyme purification.** Ten cerebella taken from Sprague Dawley rats (male, 250-350 g) were homogenized in 50 ml of ice-cold buffer A [10 mM Tris-HCl (pH 7.5) containing 1 mM DTT, 1 mM EDTA], and all subsequent purification procedures were carried out at 4 °C. The homogenate was centrifuged at 100,000  $\times$  g for 120 min, and the supernatant was loaded onto a 25 ml DEAE-agarose column equilibrated with buffer A. The column was washed with 50 ml buffer A and eluted with a 200 ml linear gradient of 0-500 mM NaCl in buffer A. Fractions (4 ml) were collected and assayed for enzyme activity. Fractions representing the active peak for BK from the DEAE-agarose column were pooled and concentrated to 15 ml by membrane ultrafiltration, and loaded onto a 5 ml, 2',5'-ADP-agarose column equilibrated with buffer B [10 mM Tris-HCl (pH 7.0) containing 1 mM DTT, 1 mM EDTA, and 10% glycerol]. The column was subsequently washed with 20 ml of buffer B, 10 ml of 0.5 M NaCl in buffer B, 10 ml of 0.5 mM NADH in buffer B, 10 ml of 0.5 mM NADP<sup>+</sup> in buffer B, and 20 ml of buffer B. The NOS activity was finally eluted with 10 ml of 10 mM NADPH in buffer B. The affinity eluate was concentrated to 1.5 ml and loaded onto a 5 ml DEAE-agarose column equilibrated with buffer B. The column was washed with 20 ml buffer B and eluted with a 100 ml linear gradient of 0-300 mM NaCl in buffer B. Fractions (2 ml) were collected and assayed for enzyme activity, and active fractions (8 ml) were pooled and concentrated to 0.2-0.5 ml with an Amicon PM-30 membrane.

**Determinations of molecular weight.** The molecular weight of denatured purified enzyme was determined by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE). The native molecular weight of the purified enzyme was estimated by analytical gel filtration chromatography carried out using an Alltech Macrosphere 150 column (7  $\mu$ m, 4.6  $\times$  25 cm) and a mobile phase of 10 mM Tris-HCl buffer (pH 7.5) containing 1 mM DTT and 0.2 M NaCl which was used to equilibrate the column and to elute protein samples at a flow rate of 0.5 ml/min. Eluting protein peaks (retention volumes, R<sub>v</sub>) were monitored at 280 nm. Standard proteins (M<sub>r</sub>) used were bovine thyroglobulin (669,000, R<sub>v</sub> 1.83 ml), horse spleen apoferritin (443,000, R<sub>v</sub> 1.97 ml), sweet potato  $\beta$ -amylase (200,000, R<sub>v</sub> 2.28 ml), yeast alcohol dehydrogenase (150,000, R<sub>v</sub> 2.47 ml), and bovine serum albumin (66,000, R<sub>v</sub> 2.78 ml). The peak (R<sub>v</sub> 2.25 ml) corresponding to the eluted enzyme (230,000) was collected and assayed for NOS activity.

**Kinetic determinations.** Kinetic experiments were conducted using the standard enzyme assay described above. To determine K<sub>m</sub> values for BK, Cit<sup>1</sup>-BK and L-arginine, substrate concentrations used were 3.125, 6.25, 12.5, 25 and 50  $\mu$ M. L-NMA and L-NNA with concentrations of 0, 3.125, 6.25, 12.5, 25 and 50  $\mu$ M were used to measure K<sub>i</sub> values in the presence of above concentrations of BK. N-adamantaneacetyl-[D-Arg<sup>0</sup>, Hyp<sup>3</sup>, Thi<sup>2,4</sup>, D-Phe<sup>7</sup>]-BK with concentrations of 0, 0.625, 1.25, 2.5, 5 and 10  $\mu$ M was used to determine its K<sub>i</sub> value in the presence of the concentrations of BK given above. Kinetic data were calculated by fitting experimental data to EZ-FIT program (12).

**Isolation of enzymatic products by high performance liquid chromatography (HPLC).** Enzymatic reaction mixtures contained 0.5  $\mu$ g purified enzyme, 50 mM Tris-HCl buffer (pH 7.5), 100  $\mu$ M NADPH, 20  $\mu$ M FAD, 20  $\mu$ M FMN, 1 mM  $\text{CaCl}_2$ , 50  $\mu$ M tetrahydrobiopterin, and 100  $\mu$ M substrate in final volumes of 0.5 ml. Reaction mixtures were

TABLE I  
Purification of nNOS-II from Rat Cerebellum<sup>a</sup>

Step	Total protein (μg)	Total activity (nmol/min)	Specific activity (nmol/min/mg protein)	Recovery (%)	Purification factor (fold)
Cytosol	356,640	44.18	0.19	110	1
DEAE-agarose	69,200	27.20	0.39	61.6	2
ADP-agarose	66	19.10	289.1	43.2	1,522
DEAE-agarose	4	4.64	1,208.3	10.5	6,360

<sup>a</sup> Values of final specific activities, recovery, and fold-purification were the means of three enzyme preparations. Enzyme activity was measured spectrophotometrically. BK (100 μM) was used for enzyme assay during enzyme purification, and enzymatic reactions were initiated by the addition of protein. Protein was determined with the Bradford protein Assay.

incubated for 120 min at 37 °C, and transferred to microconcentrators (Mr cut-off 10,000, Bio-Rad) and centrifuged to remove the enzyme. Fractions through the concentrator membrane were subjected to HPLC analyses. HPLC was performed with a Shimadzu LC-10AD pump and a SPD-M6A photodiode array UV-Vis detector set at 214-219 nm. Samples of 15 μl were resolved on a μBondapak C18-column (Waters; 10 μm; 3.9 × 300 mm, inside diameter) preceded by a guard column (3.9 × 20 mm) at a flow rate of 1 ml/min. The mobile phase consisted of mixtures of: A, 0.1% trifluoroacetic acid in water; B, 0.095% trifluoroacetic acid in acetonitrile. Elution was achieved with the following gradients: 0-15 min, 0-15% linear gradient of B; 15-35 min, 15-20% linear gradient of B; and 35-40 min, 20-0% linear gradient of B. Retention times for BK, Cit<sup>1</sup>-BK, and Cit<sup>2</sup>-BK are 22.8 min, 24.3 min, and 26.3 min, respectively. For amino acid sequencing, reaction mixture samples of 200 μl were injected and peaks at 26.3 min were collected and concentrated. Incubations without substrate and without purified enzyme were used and analyzed as controls.

**Amino acid sequencing.** Amino acid sequencing was determined by automated microsequencing with Edman degradation reactions on a 475A Sequencer (Applied Biosystems, Inc.) in the University of Iowa, Protein Structure Facility. The enzymatic product (2 μg) from HPLC (Fig. 3) was sequenced in duplicate to confirm the peptide sequence.

## RESULTS AND DISCUSSION

Crude supernatant preparations from rat cerebellum contained calmodulin-independent, NOS activity capable of catalyzing the oxidation of BK to produce NO. Since the first step in

purifying arginine-utilizing neuronal nitric oxide synthase (nNOS) from rat cerebellum (13) used weak anion exchange chromatography (DEAE-agarose), this approach was taken to partially purify the BK-utilizing NOS enzyme. Elution profiles similar to those observed by Bredt and Snyder (13) reproducibly gave two calmodulin-dependent NOS activity peaks with L-arginine as substrate. Only one of these (Fig. 1, fractions 15-21) displayed calmodulin-independent NOS activity with BK as substrate. The calmodulin-independent and BK active peak was further subjected to 2',5'-ADP-agarose affinity chromatography and a second DEAE-agarose chromatographic step to afford a 6,360-fold purified enzyme preparation (Table 1). The new constitutive neuronal NOS, designated as nNOS-II to differentiate it from previously described nNOS (13), has a molecular mass of 105 KD by SDS-PAGE (Fig. 2). The apparent native molecular mass of the enzyme was 230 kD by gel filtration, indicating that nNOS-II is a homodimeric protein. Like previously characterized nNOS from cerebellum, nNOS-II requires NADPH, FAD, FMN,  $\text{Ca}^{2+}$ , and  $\text{BH}_4$  with BK as substrate. nNOS-II has stabilities similar to nNOS. nNOS-II also oxidizes L-arginine, but only when calmodulin is added along with the other cofactors.

Since BK contains both N- and C-terminal arginines, it was possible that BK could give rise to NO plus three different nonapeptide products including:  $\text{Cit}^1\text{-BK}$ ,  $\text{Cit}^9\text{-BK}$  and  $\text{Cit}^1\text{,Cit}^9\text{-BK}$ . HPLC and amino acid sequencing were used to isolate and identify the nonapeptide product formed when nNOS-II oxidized BK. The only peptide product obtained during the nNOS-II oxidation of BK was  $\text{Cit}^1\text{,Cit}^9\text{-BK}$ , where both N- and C-terminal BK-arginines were converted to their corresponding citrullines (Fig. 3A). The synthetic nonapeptide  $\text{Cit}^1\text{-BK}$  in which N-terminal arginine was replaced with citrulline was also a substrate for nNOS-II, and it too gave  $\text{Cit}^1\text{,Cit}^9\text{-BK}$  (Fig. 3B). Difficulties encountered in synthesizing  $\text{Cit}^9\text{-BK}$  precluded its use in these studies. Apparent  $K_m$  values for BK and  $\text{Cit}^1\text{-BK}$  were 8.5 and 6.2  $\mu\text{M}$ , respectively while apparent  $V_{\text{max}}$  values for BK and  $\text{Cit}^1\text{-BK}$  were 1.2 and 1.6  $\mu\text{mol/min/mg}$  protein, respectively. The kinetic results suggest that neither  $\text{Cit}^1\text{-BK}$  nor  $\text{Cit}^9\text{-BK}$  are likely to accumulate as products during the course of nNOS-II oxidations of BK. The apparent  $K_m$  of nNOS-II for L-arginine was 10.6  $\mu\text{M}$ , slightly higher than the reported value of 2  $\mu\text{M}$  for previously described nNOS (13). With L-arginine as substrate nNOS-II and nNOS display apparent  $V_{\text{max}}$  values of 0.85 and 0.96  $\mu\text{mol/min/mg}$  protein, respectively.

With nNOS-II and BK as substrate, typical NOS inhibitors (1-4), L-NMA and L-NNA, were reversible, competitive inhibitors with apparent  $K_i$  values of 8.6  $\mu\text{M}$  and 23.8  $\mu\text{M}$  respectively. These  $K_i$  values are significantly different than previously established  $K_i$  values for L-NMA of 1.4  $\mu\text{M}$  and L-NNA of 4.4  $\mu\text{M}$  with L-arginine as substrate for nNOS (1-4). Known potent specific B2 receptor antagonists [*N*-adamantaneacetyl-D-Arg<sup>0</sup>, Hyp<sup>3</sup>, Thi<sup>5,8</sup>, D-Phe<sup>7</sup>]-BK and [Thi<sup>5,8</sup>, D-Phe<sup>7</sup>]-BK (14-16) were examined for their possible effects vs. nNOS-II activity. While [*N*-adamantaneacetyl-D-Arg<sup>0</sup>, Hyp<sup>3</sup>, Thi<sup>5,8</sup>, D-Phe<sup>7</sup>]-BK competitively inhibited nNOS-

II activity vs. BK with an apparent  $K_i$  of  $2.5 \mu\text{M}$ ,  $[\text{Thi}^{4,8}, \text{D-Phe}^7]\text{-BK}$  had no effect on NOS-II activity over a concentration range of  $0.5 - 100 \mu\text{M}$ .

This study reports the discovery of a new constitutive neuronal NOS (nNOS-II) in rat cerebellum that directly oxidizes both L-arginine (calmodulin dependent) and oligopeptides (calmodulin independent) as sources of NO (Fig. 4). Peptide products, and some catalytic and kinetic properties were defined for nNOS-II. Although NO is apparently formed from dipeptides like Arg-Arg and Arg-Phe by crude enzyme preparations from cultured endothelial and macrophage cells (17-19), this is the first report describing the oxidation of oligopeptides by a mammalian NOS, and the inhibition of an NOS by peptide antagonists. Our results suggest that natural endogenous peptide substrates other than BK may exist for nNOS-II, and that oligopeptide utilizing endothelial and macrophage NOSs remain to be identified. The broad implications of this work are that arginine-rich peptides of greater or lesser size than BK can serve as NOS substrates to form NO, and that natural or synthetic peptide antagonists can function as NOS inhibitors.

### ACKNOWLEDGMENTS

The authors thank Professors Robert J. Linhardt and Michael W. Duffel for their valuable comments and, and YC thanks the Center for Biocatalysis and Bioprocessing for fellowship support.

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